**Introduction**

Now here is a blog post that has been sitting on the shelf far longer than it should have. Over a year ago I was having when estimating the parameters of a GARCH(1,1) model in R. I documented the behavior of parameter estimates (with a focus on \beta) and perceived pathological behavior when those estimates are computed using **fGarch**. I called for help from the R community

I was not disappointed in the feedback., and there were some comments on Reddit that were helpful, but I think the best feedback I got was through my own e-mail.

I look into two more modern packages for GARCH modelling, [**rugarch**](https://cran.r-project.org/web/packages/rugarch/index.html) (for univariate GARCH models) and [**rmgarch**](https://cran.r-project.org/web/packages/rmgarch/index.html) (for multivariate GARCH models). I had not heard of these packages before, so I’m very thankful for the suggestion. Since I’m interested in univariate time series for now, I only looked at **rugarch**. The package appears to have more features and power than **fGarch**, which may explain why it seems more difficult to use.

The ease of getting daily data (particularly for USD/JPY around the time of Asian financial crises, which was an intended application of a test statistic I’m studying) motivated my interest in daily data. His comments, though, may lead me to reconsider this application. (I might try to detect the 2010 eurozone financial crises via EUR/USD instead. I can get free intraday data from [HistData.com](http://www.histdata.com/download-free-forex-data/) for this.) However, if standard error estimates cannot be trusted for small sample sizes, our test statistic would still be in trouble since it involves estimating parameters even for small sample sizes.

That may be true, but simulated data is important since it can be considered a statistician’s best-case scenario. Additionally, the properties of the process that generated simulated data are known *a priori*, including the values of the generating parameters and whether certain hypotheses (such as whether there is a structural change in the series) are true. This allows for sanity checks of estimators and tests. This is impossible for real-world since we don’t have the *a priori* knowledge needed.

I repeat the simulations but with \alpha  0.6since these values are supposedly more common than my choice of \alpha = \beta = 0.2. It’s a good suggestion and I will consider parameters in this range in addition to \alpha = \beta = 0.2in this post. However, my simulations seemed to suggest that when \alpha = \beta = 0.2, the estimation procedures nevertheless seem to want to be near the range of large \beta. I’m also surprised since my advisor gave me the impression that GARCH processes with either \alphaor \betalarge are more difficult to work with. Finally, if the estimators are strongly biased, we might expect to see most estimated parameters to lie in that range, though that does not mean the “correct” values lie in that range. My simulations suggest **fGarch** struggles to discover \alpha = \beta = 0.2even when those parameters are “true.”” Prof. Santos’ comment leads me to desire a metastudy about what common estimates of GARCH parameters are on real world. (There may or may not be one; I haven’t checked. If anyone knows of one, please share.)

My advisor contacted another expert on GARCH models and got some feedback. Supposedly the standard error for \betais large, so there should be great variation in parameter estimates. Some of my simulations agreed with this behavior even for small sample sizes, but at the same time showed an uncomfortable bias towards \beta = 0and \beta = 1. This might be a consequence of the optimization procedures, as I hypothesized.

So given this feedback, I will be conducting more simulation experiments. I won’t be looking at **fGarch** or **tseries** anymore; I will be working exclusively with **rugarch**. I will explore different optimization procedures supported by the package. I won’t be creating plots like I did in my first post; those plots were meant only to show the existence of a problem and its severity. Instead I will be looking at properties of the resulting estimators produced by different optimization procedures.

**Introduction to rugarch**

As mentioned above, **rugarch** is a package for working with GARCH models; a major use case is estimating their parameters, obviously. Here I will demonstrate how to specify a GARCH model, simulate data from the model, and estimate parameters. After this we can dive into simulation studies.

library(rugarch)

## Loading required package: parallel

##

## Attaching package: 'rugarch'

## The following object is masked from 'package:stats':

##

## sigma

**Specifying a \text{GARCH}(1, 1)model**

To work with a GARCH model we need to specify it. The function for doing this is ugarchspec(). I think the parameters variance.model and mean.model are the most important parameters.

variance.model is a list with named entries, perhaps the two most interesting being model and garchOrder. model is a string specify which type of GARCH model is being fitted. Many major classes of GARCH models (such as EGARCH, IGARCH, etc.) are supported; for the “vanilla” GARCH model, set this to "sGARCH" (or just omit it; the standard model is the default). garchOrder is a vector for the order of the ARCH and GARCH components of the model.

mean.model allows for fitting ARMA-GARCH models, and functions like variance.model in that it accepts a list of named entries, the most interesting being armaOrder and include.mean. armaOrder is like garchOrder; it’s a vector specifying the order of the ARMA model. include.mean is a boolean that, if true, allows for the ARMA part of the model to have non-zero mean.

When simulating a process, we need to set the values of our parameters. This is done via the fixed.pars parameter, which accepts a list of named elements, the elements of the list being numeric. They need to fit the conventions the function uses for parameters; for example, if we want to set the parameters of a \text{GARCH}(1,1)model, the names of our list elements should be "alpha1" and "beta1". If the plan is to simulate a model, every parameter in the model should be set this way.

There are other parameters interesting in their own right but I focus on these since the default specification is an ARMA-GARCH model with ARMA order of (1,1)with non-zero mean and a GARCH model of order (1,1). This is not a vanilla \text{GARCH}(1,1)model as I desire, so I almost always change this.

spec1 <- ugarchspec(mean.model = list(armaOrder = c(0,0), include.mean = FALSE),

fixed.pars = list("omega" = 0.2, "alpha1" = 0.2,

"beta1" = 0.2))

spec2 <- ugarchspec(mean.model = list(armaOrder = c(0,0), include.mean = FALSE),

fixed.pars = list("omega" = 0.2, "alpha1" = 0.1,

"beta1" = 0.7))

show(spec1)

##

## \*---------------------------------\*

## \* GARCH Model Spec \*

## \*---------------------------------\*

##

## Conditional Variance Dynamics

## ------------------------------------

## GARCH Model : sGARCH(1,1)

## Variance Targeting : FALSE

##

## Conditional Mean Dynamics

## ------------------------------------

## Mean Model : ARFIMA(0,0,0)

## Include Mean : FALSE

## GARCH-in-Mean : FALSE

##

## Conditional Distribution

## ------------------------------------

## Distribution : norm

## Includes Skew : FALSE

## Includes Shape : FALSE

## Includes Lambda : FALSE

show(spec2)

##

## \*---------------------------------\*

## \* GARCH Model Spec \*

## \*---------------------------------\*

##

## Conditional Variance Dynamics

## ------------------------------------

## GARCH Model : sGARCH(1,1)

## Variance Targeting : FALSE

##

## Conditional Mean Dynamics

## ------------------------------------

## Mean Model : ARFIMA(0,0,0)

## Include Mean : FALSE

## GARCH-in-Mean : FALSE

##

## Conditional Distribution

## ------------------------------------

## Distribution : norm

## Includes Skew : FALSE

## Includes Shape : FALSE

## Includes Lambda : FALSE

**Simulating a GARCH process**

The function ugarchpath() simulates GARCH models specified via ugarchspec(). The function needs a specification objectect created by ugarchspec() first. The parameters n.sim and n.start specify the size of the process and the length of the burn-in period, respectively (with defaults 1000 and 0, respectively; I strongly recommend setting the burn-in period to at least 500, but I go for 1000). The function creates an object that contains not only the simulated series but also residuals and \sigma_t.

The rseed parameter controls the random seed the function uses for generating data. Be warned that set.seed() is effectively ignored by this function, so if you want consistent results, you will need to set this parameter.

The plot() method accompanying these objects is not completely transparent; there are a few plots it could create and when calling plot() on a uGARCHpath object in the command line users are prompted to input a number corresponding to the desired plot. This is a pain sometimes so don’t forget to pass the desired plot’s number to the which parameter to avoid the prompt; setting which = 2 will give the plot of the series proper.

old\_par <- par()

par(mfrow = c(2, 2))

x\_obj <- ugarchpath(spec1, n.sim = 1000, n.start = 1000, rseed = 111217)

show(x\_obj)

##

## \*------------------------------------\*

## \* GARCH Model Path Simulation \*

## \*------------------------------------\*

## Model: sGARCH

## Horizon: 1000

## Simulations: 1

## Seed Sigma2.Mean Sigma2.Min Sigma2.Max Series.Mean

## sim1 111217 0.332 0.251 0.915 0.000165

## Mean(All) 0 0.332 0.251 0.915 0.000165

## Unconditional NA 0.333 NA NA 0.000000

## Series.Min Series.Max

## sim1 -1.76 1.62

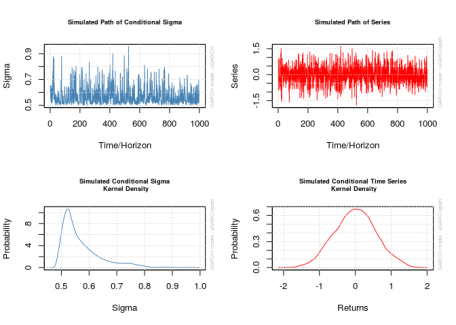
## Mean(All) -1.76 1.62

## Unconditional NA NA

for (i in 1:4) {

plot(x\_obj, which = i)

}



par(old\_par)

## Warning in par(old\_par): graphical parameter "cin" cannot be set

## Warning in par(old\_par): graphical parameter "cra" cannot be set

## Warning in par(old\_par): graphical parameter "csi" cannot be set

## Warning in par(old\_par): graphical parameter "cxy" cannot be set

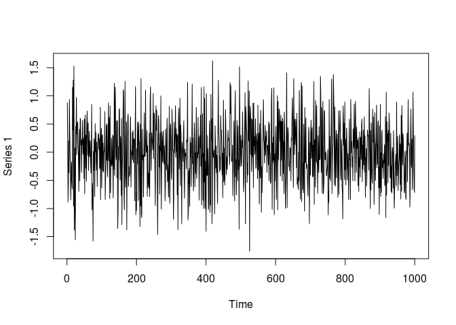
## Warning in par(old\_par): graphical parameter "din" cannot be set

## Warning in par(old\_par): graphical parameter "page" cannot be set

# The actual series

x1 <- x\_obj@path$seriesSim

plot.ts(x1)



**Fitting a \text{GARCH}(1,1)model**

The ugarchfit() function fits GARCH models. The function needs a specification and a dataset. The solver parameter accepts a string stating which numerical optimizer to use to find the parameter estimates. Most of the parameters of the function manage interfacing with the numerical optimizer. In particular, solver.control can be given a list of arguments to pass to the optimizer. We will be looking at this in more detail later.

The specification used for generating the simulated data won’t be appropriate for ugarchfit(), since it contains fixed values for its parameters. In my case I will need to create a second specification object.

spec <- ugarchspec(mean.model = list(armaOrder = c(0, 0), include.mean = FALSE))

fit <- ugarchfit(spec, data = x1)

show(fit)

##

## \*---------------------------------\*

## \* GARCH Model Fit \*

## \*---------------------------------\*

##

## Conditional Variance Dynamics

## -----------------------------------

## GARCH Model : sGARCH(1,1)

## Mean Model : ARFIMA(0,0,0)

## Distribution : norm

##

## Optimal Parameters

## ------------------------------------

## Estimate Std. Error t value Pr(>|t|)

## omega 0.000713 0.001258 0.56696 0.57074

## alpha1 0.002905 0.003714 0.78206 0.43418

## beta1 0.994744 0.000357 2786.08631 0.00000

##

## Robust Standard Errors:

## Estimate Std. Error t value Pr(>|t|)

## omega 0.000713 0.001217 0.58597 0.55789

## alpha1 0.002905 0.003661 0.79330 0.42760

## beta1 0.994744 0.000137 7250.45186 0.00000

##

## LogLikelihood : -860.486

##

## Information Criteria

## ------------------------------------

##

## Akaike 1.7270

## Bayes 1.7417

## Shibata 1.7270

## Hannan-Quinn 1.7326

##

## Weighted Ljung-Box Test on Standardized Residuals

## ------------------------------------

## statistic p-value

## Lag[1] 3.998 0.04555

## Lag[2\*(p+q)+(p+q)-1][2] 4.507 0.05511

## Lag[4\*(p+q)+(p+q)-1][5] 9.108 0.01555

## d.o.f=0

## H0 : No serial correlation

##

## Weighted Ljung-Box Test on Standardized Squared Residuals

## ------------------------------------

## statistic p-value

## Lag[1] 29.12 6.786e-08

## Lag[2\*(p+q)+(p+q)-1][5] 31.03 1.621e-08

## Lag[4\*(p+q)+(p+q)-1][9] 32.26 1.044e-07

## d.o.f=2

##

## Weighted ARCH LM Tests

## ------------------------------------

## Statistic Shape Scale P-Value

## ARCH Lag[3] 1.422 0.500 2.000 0.2331

## ARCH Lag[5] 2.407 1.440 1.667 0.3882

## ARCH Lag[7] 2.627 2.315 1.543 0.5865

##

## Nyblom stability test

## ------------------------------------

## Joint Statistic: 0.9518

## Individual Statistics:

## omega 0.3296

## alpha1 0.2880

## beta1 0.3195

##

## Asymptotic Critical Values (10% 5% 1%)

## Joint Statistic: 0.846 1.01 1.35

## Individual Statistic: 0.35 0.47 0.75

##

## Sign Bias Test

## ------------------------------------

## t-value prob sig

## Sign Bias 0.3946 6.933e-01

## Negative Sign Bias 3.2332 1.264e-03 \*\*\*

## Positive Sign Bias 4.2142 2.734e-05 \*\*\*

## Joint Effect 28.2986 3.144e-06 \*\*\*

##

##

## Adjusted Pearson Goodness-of-Fit Test:

## ------------------------------------

## group statistic p-value(g-1)

## 1 20 20.28 0.3779

## 2 30 26.54 0.5965

## 3 40 36.56 0.5817

## 4 50 47.10 0.5505

##

##

## Elapsed time : 2.60606

par(mfrow = c(3, 4))

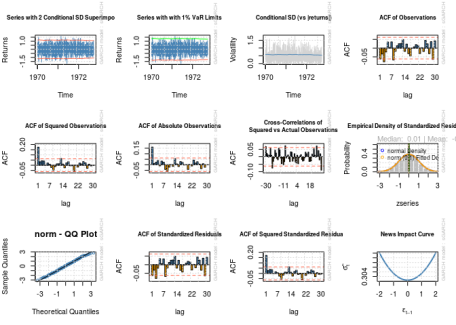
for (i in 1:12) {

plot(fit, which = i)

}

##

## please wait...calculating quantiles...



par(old\_par)

## Warning in par(old\_par): graphical parameter "cin" cannot be set

## Warning in par(old\_par): graphical parameter "cra" cannot be set

## Warning in par(old\_par): graphical parameter "csi" cannot be set

## Warning in par(old\_par): graphical parameter "cxy" cannot be set

## Warning in par(old\_par): graphical parameter "din" cannot be set

## Warning in par(old\_par): graphical parameter "page" cannot be set

Notice the estimated parameters and standard errors? The estimates are nowhere near the “correct” numbers even for a sample size of 1000, and there is no way a reasonable confidence interval based on the estimated standard errors would contain the correct values. It looks like the problems I documented in my last post have not gone away.

Out of curiosity, what would happen with the other specification, one in the range Prof. Santos suggested?

x\_obj <- ugarchpath(spec2, n.start = 1000, rseed = 111317)

x2 <- x\_obj@path$seriesSim

fit <- ugarchfit(spec, x2)

show(fit)

##

## \*---------------------------------\*

## \* GARCH Model Fit \*

## \*---------------------------------\*

##

## Conditional Variance Dynamics

## -----------------------------------

## GARCH Model : sGARCH(1,1)

## Mean Model : ARFIMA(0,0,0)

## Distribution : norm

##

## Optimal Parameters

## ------------------------------------

## Estimate Std. Error t value Pr(>|t|)

## omega 0.001076 0.002501 0.43025 0.66701

## alpha1 0.001992 0.002948 0.67573 0.49921

## beta1 0.997008 0.000472 2112.23364 0.00000

##

## Robust Standard Errors:

## Estimate Std. Error t value Pr(>|t|)

## omega 0.001076 0.002957 0.36389 0.71594

## alpha1 0.001992 0.003510 0.56767 0.57026

## beta1 0.997008 0.000359 2777.24390 0.00000

##

## LogLikelihood : -1375.951

##

## Information Criteria

## ------------------------------------

##

## Akaike 2.7579

## Bayes 2.7726

## Shibata 2.7579

## Hannan-Quinn 2.7635

##

## Weighted Ljung-Box Test on Standardized Residuals

## ------------------------------------

## statistic p-value

## Lag[1] 0.9901 0.3197

## Lag[2\*(p+q)+(p+q)-1][2] 1.0274 0.4894

## Lag[4\*(p+q)+(p+q)-1][5] 3.4159 0.3363

## d.o.f=0

## H0 : No serial correlation

##

## Weighted Ljung-Box Test on Standardized Squared Residuals

## ------------------------------------

## statistic p-value

## Lag[1] 3.768 0.05226

## Lag[2\*(p+q)+(p+q)-1][5] 4.986 0.15424

## Lag[4\*(p+q)+(p+q)-1][9] 7.473 0.16272

## d.o.f=2

##

## Weighted ARCH LM Tests

## ------------------------------------

## Statistic Shape Scale P-Value

## ARCH Lag[3] 0.2232 0.500 2.000 0.6366

## ARCH Lag[5] 0.4793 1.440 1.667 0.8897

## ARCH Lag[7] 2.2303 2.315 1.543 0.6686

##

## Nyblom stability test

## ------------------------------------

## Joint Statistic: 0.3868

## Individual Statistics:

## omega 0.2682

## alpha1 0.2683

## beta1 0.2669

##

## Asymptotic Critical Values (10% 5% 1%)

## Joint Statistic: 0.846 1.01 1.35

## Individual Statistic: 0.35 0.47 0.75

##

## Sign Bias Test

## ------------------------------------

## t-value prob sig

## Sign Bias 0.5793 0.5625

## Negative Sign Bias 1.3358 0.1819

## Positive Sign Bias 1.5552 0.1202

## Joint Effect 5.3837 0.1458

##

##

## Adjusted Pearson Goodness-of-Fit Test:

## ------------------------------------

## group statistic p-value(g-1)

## 1 20 24.24 0.1871

## 2 30 30.50 0.3894

## 3 40 38.88 0.4753

## 4 50 48.40 0.4974

##

##

## Elapsed time : 2.841597

That’s no better Now let’s see what happens when we use different optimization routines.

**Optimization and Parameter Estimation in rugarch**

**Choice of optimizer**

ugarchfit()‘s default parameters did a good job of finding appropriate parameters for what I will refer to as model 2 (where \alpha = 0.1and \beta = 0.7) but not for model 1 (\alpha = \beta = 0.2). What I want to know is when one solver seems to beat another.

Below is a helper function that simplifies the process of fitting a GARCH model’s parameters and extracting the log-likelihood, parameter values, and standard errors while allowing for different values to be passed to solver and solver.control.

evalSolverFit <- function(spec, data, solver = "solnp",

solver.control = list()) {

# Calls ugarchfit(spec, data, solver, solver.control), and returns a vector

# containing the log likelihood, parameters, and parameter standard errors.

# Parameters are equivalent to those seen in ugarchfit(). If the solver fails

# to converge, NA will be returned

vec <- NA

tryCatch({

fit <- ugarchfit(spec = spec, data = data, solver = solver,

solver.control = solver.control)

coef\_se\_names <- paste("se", names(fit@fit$coef), sep = ".")

se <- fit@fit$se.coef

names(se) <- coef\_se\_names

robust\_coef\_se\_names <- paste("robust.se", names(fit@fit$coef), sep = ".")

robust.se <- fit@fit$robust.se.coef

names(robust.se) <- robust\_coef\_se\_names

vec <- c(fit@fit$coef, se, robust.se)

vec["LLH"] <- fit@fit$LLH

}, error = function(w) {

NA

})

return(vec)

}

Below I list out all optimization schemes I will consider. I only fiddle with solver.control, but there may be other parameters that could help the numerical optimization routines, namely numderiv.control, which are control arguments passed to the numerical routines responsible for standard error computation. This utilizes the package **numDeriv** which performs numerical differentiation.

solvers <- list( # A list of lists where each sublist contains parameters to

# pass to a solver

list("solver" = "nlminb", "solver.control" = list()),

list("solver" = "solnp", "solver.control" = list()),

list("solver" = "lbfgs", "solver.control" = list()),

list("solver" = "gosolnp", "solver.control" = list(

"n.restarts" = 100, "n.sim" = 100

)),

list("solver" = "hybrid", "solver.control" = list()),

list("solver" = "nloptr", "solver.control" = list("solver" = 1)), # COBYLA

list("solver" = "nloptr", "solver.control" = list("solver" = 2)), # BOBYQA

list("solver" = "nloptr", "solver.control" = list("solver" = 3)), # PRAXIS

list("solver" = "nloptr",

"solver.control" = list("solver" = 4)), # NELDERMEAD

list("solver" = "nloptr", "solver.control" = list("solver" = 5)), # SBPLX

list("solver" = "nloptr",

"solver.control" = list("solver" = 6)), # AUGLAG+COBYLA

list("solver" = "nloptr",

"solver.control" = list("solver" = 7)), # AUGLAG+BOBYQA

list("solver" = "nloptr",

"solver.control" = list("solver" = 8)), # AUGLAG+PRAXIS

list("solver" = "nloptr",

"solver.control" = list("solver" = 9)), # AUGLAG+NELDERMEAD

list("solver" = "nloptr",

"solver.control" = list("solver" = 10)) # AUGLAG+SBPLX

)

tags <- c( # Names for the above list

"nlminb",

"solnp",

"lbfgs",

"gosolnp",

"hybrid",

"nloptr+COBYLA",

"nloptr+BOBYQA",

"nloptr+PRAXIS",

"nloptr+NELDERMEAD",

"nloptr+SBPLX",

"nloptr+AUGLAG+COBYLA",

"nloptr+AUGLAG+BOBYQA",

"nloptr+AUGLAG+PRAXIS",

"nloptr+AUGLAG+NELDERMEAD",

"nloptr+AUGLAG+SBPLX"

)

names(solvers) <- tags

Now let’s run the gauntlet of optimization choices and see which produces the estimates with the largest log likelihood for data generated by model 1. The lbfgs method unfortunately does not converge for this series, so I omit it.

optMethodCompare <- function(data, spec, solvers) {

# Runs all solvers in a list for a dataset

#

# Args:

# data: An object to pass to ugarchfit's data parameter containing the data

# to fit

# spec: A specification created by ugarchspec to pass to ugarchfit

# solvers: A list of lists containing strings of solvers and a list for

# solver.control

#

# Return:

# A matrix containing the result of the solvers (including parameters, se's,

# and LLH)

model\_solutions <- lapply(solvers, function(s) {

args <- s

args[["spec"]] <- spec

args[["data"]] <- data

res <- do.call(evalSolverFit, args = args)

return(res)

})

model\_solutions <- do.call(rbind, model\_solutions)

return(model\_solutions)

}

round(optMethodCompare(x1, spec, solvers[c(1:2, 4:15)]), digits = 4)

## omega alpha1 beta1 se.omega se.alpha1 se.beta1 robust.se.omega robust.se.alpha1 robust.se.beta1 LLH

## ------------------------- ------- ------- ------- --------- ---------- --------- ---------------- ----------------- ---------------- ----------

## nlminb 0.2689 0.1774 0.0000 0.0787 0.0472 0.2447 0.0890 0.0352 0.2830 -849.6927

## solnp 0.0007 0.0029 0.9947 0.0013 0.0037 0.0004 0.0012 0.0037 0.0001 -860.4860

## gosolnp 0.2689 0.1774 0.0000 0.0787 0.0472 0.2446 0.0890 0.0352 0.2828 -849.6927

## hybrid 0.0007 0.0029 0.9947 0.0013 0.0037 0.0004 0.0012 0.0037 0.0001 -860.4860

## nloptr+COBYLA 0.0006 0.0899 0.9101 0.0039 0.0306 0.0370 0.0052 0.0527 0.0677 -871.5006

## nloptr+BOBYQA 0.0003 0.0907 0.9093 0.0040 0.0298 0.0375 0.0057 0.0532 0.0718 -872.3436

## nloptr+PRAXIS 0.2689 0.1774 0.0000 0.0786 0.0472 0.2444 0.0888 0.0352 0.2823 -849.6927

## nloptr+NELDERMEAD 0.0010 0.0033 0.9935 0.0013 0.0039 0.0004 0.0013 0.0038 0.0001 -860.4845

## nloptr+SBPLX 0.0010 0.1000 0.9000 0.0042 0.0324 0.0386 0.0055 0.0536 0.0680 -872.2736

## nloptr+AUGLAG+COBYLA 0.0006 0.0899 0.9101 0.0039 0.0306 0.0370 0.0052 0.0527 0.0677 -871.5006

## nloptr+AUGLAG+BOBYQA 0.0003 0.0907 0.9093 0.0040 0.0298 0.0375 0.0057 0.0532 0.0718 -872.3412

## nloptr+AUGLAG+PRAXIS 0.1246 0.1232 0.4948 0.0620 0.0475 0.2225 0.0701 0.0439 0.2508 -851.0547

## nloptr+AUGLAG+NELDERMEAD 0.2689 0.1774 0.0000 0.0786 0.0472 0.2445 0.0889 0.0352 0.2826 -849.6927

## nloptr+AUGLAG+SBPLX 0.0010 0.1000 0.9000 0.0042 0.0324 0.0386 0.0055 0.0536 0.0680 -872.2736

According the the maximum likelihood criterion, the “best” result is achieved by gosolnp. The result has the unfortunate property that \beta \approx 0, which is certainly not true, but at least the standard error for \betawould create a confidence interval that contains \beta‘s true value. Of these, my preferred estimates are produced by AUGLAG+PRAXIS, as \betaseems reasonable and in fact the estimates are all close to the truth, (at least in the sense that the confidence intervals contain the true values), but unfortunately the estimates do *not* maximize the log likelihood, even though they are the most reasonable.

If we looked at model 2, what do we see? Again, lbfgs does not converge so I omit it. Unfortunately, nlminb does not converge either, so it too must be omitted.

round(optMethodCompare(x2, spec, solvers[c(2, 4:15)]), digits = 4)

## omega alpha1 beta1 se.omega se.alpha1 se.beta1 robust.se.omega robust.se.alpha1 robust.se.beta1 LLH

## ------------------------- ------- ------- ------- --------- ---------- --------- ---------------- ----------------- ---------------- ----------

## solnp 0.0011 0.0020 0.9970 0.0025 0.0029 0.0005 0.0030 0.0035 0.0004 -1375.951

## gosolnp 0.0011 0.0020 0.9970 0.0025 0.0029 0.0005 0.0030 0.0035 0.0004 -1375.951

## hybrid 0.0011 0.0020 0.9970 0.0025 0.0029 0.0005 0.0030 0.0035 0.0004 -1375.951

## nloptr+COBYLA 0.0016 0.0888 0.9112 0.0175 0.0619 0.0790 0.0540 0.2167 0.2834 -1394.529

## nloptr+BOBYQA 0.0010 0.0892 0.9108 0.0194 0.0659 0.0874 0.0710 0.2631 0.3572 -1395.310

## nloptr+PRAXIS 0.5018 0.0739 0.3803 0.3178 0.0401 0.3637 0.2777 0.0341 0.3225 -1373.632

## nloptr+NELDERMEAD 0.0028 0.0026 0.9944 0.0028 0.0031 0.0004 0.0031 0.0035 0.0001 -1375.976

## nloptr+SBPLX 0.0029 0.1000 0.9000 0.0146 0.0475 0.0577 0.0275 0.1108 0.1408 -1395.807

## nloptr+AUGLAG+COBYLA 0.0016 0.0888 0.9112 0.0175 0.0619 0.0790 0.0540 0.2167 0.2834 -1394.529

## nloptr+AUGLAG+BOBYQA 0.0010 0.0892 0.9108 0.0194 0.0659 0.0874 0.0710 0.2631 0.3572 -1395.310

## nloptr+AUGLAG+PRAXIS 0.5018 0.0739 0.3803 0.3178 0.0401 0.3637 0.2777 0.0341 0.3225 -1373.632

## nloptr+AUGLAG+NELDERMEAD 0.0001 0.0000 1.0000 0.0003 0.0003 0.0000 0.0004 0.0004 0.0000 -1375.885

## nloptr+AUGLAG+SBPLX 0.0029 0.1000 0.9000 0.0146 0.0475 0.0577 0.0275 0.1108 0.1408 -1395.807

Here it was PRAXIS and AUGLAG+PRAXIS that gave the “optimal” result, and it was only those two methods that did. Other optimizers gave visibly bad results. That said, the “optimal” solution is the preferred on with the parameters being nonzero and their confidence intervals containing the correct values.

What happens if we restrict the sample to size 100? (lbfgs still does not work.)

round(optMethodCompare(x1[1:100], spec, solvers[c(1:2, 4:15)]), digits = 4)

## omega alpha1 beta1 se.omega se.alpha1 se.beta1 robust.se.omega robust.se.alpha1 robust.se.beta1 LLH

## ------------------------- ------- ------- ------- --------- ---------- --------- ---------------- ----------------- ---------------- ---------

## nlminb 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## solnp 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## gosolnp 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## hybrid 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## nloptr+COBYLA 0.0007 0.1202 0.8798 0.0085 0.0999 0.0983 0.0081 0.1875 0.1778 -85.3121

## nloptr+BOBYQA 0.0005 0.1190 0.8810 0.0085 0.0994 0.0992 0.0084 0.1892 0.1831 -85.3717

## nloptr+PRAXIS 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## nloptr+NELDERMEAD 0.0451 0.2742 0.5920 0.0281 0.1230 0.1297 0.0191 0.0906 0.0667 -80.6587

## nloptr+SBPLX 0.0433 0.2740 0.5998 0.0269 0.1237 0.1268 0.0182 0.0916 0.0648 -80.6616

## nloptr+AUGLAG+COBYLA 0.0007 0.1202 0.8798 0.0085 0.0999 0.0983 0.0081 0.1875 0.1778 -85.3121

## nloptr+AUGLAG+BOBYQA 0.0005 0.1190 0.8810 0.0085 0.0994 0.0992 0.0084 0.1892 0.1831 -85.3717

## nloptr+AUGLAG+PRAXIS 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## nloptr+AUGLAG+NELDERMEAD 0.0451 0.2742 0.5921 0.0280 0.1229 0.1296 0.0191 0.0905 0.0667 -80.6587

## nloptr+AUGLAG+SBPLX 0.0450 0.2742 0.5924 0.0280 0.1230 0.1295 0.0191 0.0906 0.0666 -80.6587

round(optMethodCompare(x2[1:100], spec, solvers[c(1:2, 4:15)]), digits = 4)

## omega alpha1 beta1 se.omega se.alpha1 se.beta1 robust.se.omega robust.se.alpha1 robust.se.beta1 LLH

## ------------------------- ------- ------- ------- --------- ---------- --------- ---------------- ----------------- ---------------- ----------

## nlminb 0.7592 0.0850 0.0000 2.1366 0.4813 3.0945 7.5439 1.7763 11.0570 -132.4614

## solnp 0.0008 0.0000 0.9990 0.0291 0.0417 0.0066 0.0232 0.0328 0.0034 -132.9182

## gosolnp 0.0537 0.0000 0.9369 0.0521 0.0087 0.0713 0.0430 0.0012 0.0529 -132.9124

## hybrid 0.0008 0.0000 0.9990 0.0291 0.0417 0.0066 0.0232 0.0328 0.0034 -132.9182

## nloptr+COBYLA 0.0014 0.0899 0.9101 0.0259 0.0330 0.1192 0.0709 0.0943 0.1344 -135.7495

## nloptr+BOBYQA 0.0008 0.0905 0.9095 0.0220 0.0051 0.1145 0.0687 0.0907 0.1261 -135.8228

## nloptr+PRAXIS 0.0602 0.0000 0.9293 0.0522 0.0088 0.0773 0.0462 0.0015 0.0565 -132.9125

## nloptr+NELDERMEAD 0.0024 0.0000 0.9971 0.0473 0.0629 0.0116 0.0499 0.0680 0.0066 -132.9186

## nloptr+SBPLX 0.0027 0.1000 0.9000 0.0238 0.0493 0.1308 0.0769 0.1049 0.1535 -135.9175

## nloptr+AUGLAG+COBYLA 0.0014 0.0899 0.9101 0.0259 0.0330 0.1192 0.0709 0.0943 0.1344 -135.7495

## nloptr+AUGLAG+BOBYQA 0.0008 0.0905 0.9095 0.0221 0.0053 0.1145 0.0687 0.0907 0.1262 -135.8226

## nloptr+AUGLAG+PRAXIS 0.0602 0.0000 0.9294 0.0523 0.0090 0.0771 0.0462 0.0014 0.0565 -132.9125

## nloptr+AUGLAG+NELDERMEAD 0.0000 0.0000 0.9999 0.0027 0.0006 0.0005 0.0013 0.0004 0.0003 -132.9180

## nloptr+AUGLAG+SBPLX 0.0027 0.1000 0.9000 0.0238 0.0493 0.1308 0.0769 0.1049 0.1535 -135.9175

The results are not thrilling. The “best” result for the series generated by model 1 was attained by multiple solvers, and the 95% confidence interval (CI) for \omegawould not contain \omega‘s true value, though the CIs for the other parameters would contain their true values. For the series generated by model 2 the best result was attained by the nlminb solver; the parameter values are not plausible and the standard errors are huge. At least the CI would contain the correct value.

From here we should no longer stick to two series but see the performance of these methods on many simulated series generated by both models. Simulations in this post will be too computationally intensive for my laptop so I will use my department’s supercomputer to perform them, taking advantage of its many cores for parallelization.

library(foreach)

library(doParallel)

logfile <- ""

# logfile <- "outfile.log"

# if (!file.exists(logfile)) {

# file.create(logfile)

# }

cl <- makeCluster(detectCores() - 1, outfile = logfile)

registerDoParallel(cl)

optMethodSims <- function(gen\_spec, n.sim = 1000, m.sim = 1000,

fit\_spec = ugarchspec(mean.model = list(

armaOrder = c(0,0), include.mean = FALSE)),

solvers = list("solnp" = list(

"solver" = "solnp", "solver.control" = list())),

rseed = NA, verbose = FALSE) {

# Performs simulations in parallel of GARCH processes via rugarch and returns

# a list with the results of different optimization routines

#

# Args:

# gen\_spec: The specification for generating a GARCH sequence, produced by

# ugarchspec

# n.sim: An integer denoting the length of the simulated series

# m.sim: An integer for the number of simulated sequences to generate

# fit\_spec: A ugarchspec specification for the model to fit

# solvers: A list of lists containing strings of solvers and a list for

# solver.control

# rseed: Optional seeding value(s) for the random number generator. For

# m.sim>1, it is possible to provide either a single seed to

# initialize all values, or one seed per separate simulation (i.e.

# m.sim seeds). However, in the latter case this may result in some

# slight overhead depending on how large m.sim is

# verbose: Boolean for whether to write data tracking the progress of the

# loop into an output file

# outfile: A string for the file to store verbose output to (relevant only

# if verbose is TRUE)

#

# Return:

# A list containing the result of calling optMethodCompare on each generated

# sequence

fits <- foreach(i = 1:m.sim,

.packages = c("rugarch"),

.export = c("optMethodCompare", "evalSolverFit")) %dopar% {

if (is.na(rseed)) {

newseed <- NA

} else if (is.vector(rseed)) {

newseed <- rseed[i]

} else {

newseed <- rseed + i - 1

}

if (verbose) {

cat(as.character(Sys.time()), ": Now on simulation ", i, "\n")

}

sim <- ugarchpath(gen\_spec, n.sim = n.sim, n.start = 1000,

m.sim = 1, rseed = newseed)

x <- sim@path$seriesSim

optMethodCompare(x, spec = fit\_spec, solvers = solvers)

}

return(fits)

}

# Specification 1 first

spec1\_n100 <- optMethodSims(spec1, n.sim = 100, m.sim = 1000,

solvers = solvers, verbose = TRUE)

spec1\_n500 <- optMethodSims(spec1, n.sim = 500, m.sim = 1000,

solvers = solvers, verbose = TRUE)

spec1\_n1000 <- optMethodSims(spec1, n.sim = 1000, m.sim = 1000,

solvers = solvers, verbose = TRUE)

# Specification 2 next

spec2\_n100 <- optMethodSims(spec2, n.sim = 100, m.sim = 1000,

solvers = solvers, verbose = TRUE)

spec2\_n500 <- optMethodSims(spec2, n.sim = 500, m.sim = 1000,

solvers = solvers, verbose = TRUE)

spec2\_n1000 <- optMethodSims(spec2, n.sim = 1000, m.sim = 1000,

solvers = solvers, verbose = TRUE)

Below is a set of helper functions I will use for the analytics I want.

optMethodSims\_getAllVals <- function(param, solver, reslist) {

# Get all values for a parameter obtained by a certain solver after getting a

# list of results via optMethodSims

#

# Args:

# param: A string for the parameter to get (such as "beta1")

# solver: A string for the solver for which to get the parameter (such as

# "nlminb")

# reslist: A list created by optMethodSims

#

# Return:

# A vector of values of the parameter for each simulation

res <- sapply(reslist, function(l) {

return(l[solver, param])

})

return(res)

}

optMethodSims\_getBestVals <- function(reslist, opt\_vec = TRUE,

reslike = FALSE) {

# A function that gets the optimizer that maximized the likelihood function

# for each entry in reslist

#

# Args:

# reslist: A list created by optMethodSims

# opt\_vec: A boolean indicating whether to return a vector with the name of

# the optimizers that maximized the log likelihood

# reslike: A bookean indicating whether the resulting list should consist of

# matrices of only one row labeled "best" with a structure like

# reslist

#

# Return:

# If opt\_vec is TRUE, a list of lists, where each sublist contains a vector

# of strings naming the opimizers that maximized the likelihood function and

# a matrix of the parameters found. Otherwise, just the matrix (resembles

# the list generated by optMethodSims)

res <- lapply(reslist, function(l) {

max\_llh <- max(l[, "LLH"], na.rm = TRUE)

best\_idx <- (l[, "LLH"] == max\_llh) & (!is.na(l[, "LLH"]))

best\_mat <- l[best\_idx, , drop = FALSE]

if (opt\_vec) {

return(list("solvers" = rownames(best\_mat), "params" = best\_mat))

} else {

return(best\_mat)

}

})

if (reslike) {

res <- lapply(res, function(l) {

mat <- l$params[1, , drop = FALSE]

rownames(mat) <- "best"

return(mat)

})

}

return(res)

}

optMethodSims\_getCaptureRate <- function(param, solver, reslist, multiplier = 2,

spec, use\_robust = TRUE) {

# Gets the rate a confidence interval for a parameter captures the true value

#

# Args:

# param: A string for the parameter being worked with

# solver: A string for the solver used to estimate the parameter

# reslist: A list created by optMethodSims

# multiplier: A floating-point number for the multiplier to the standard

# error, appropriate for the desired confidence level

# spec: A ugarchspec specification with the fixed parameters containing the

# true parameter value

# use\_robust: Use robust standard errors for computing CIs

#

# Return:

# A float for the proportion of times the confidence interval captured the

# true parameter value

se\_string <- ifelse(use\_robust, "robust.se.", "se.")

est <- optMethodSims\_getAllVals(param, solver, reslist)

moe\_est <- multiplier \* optMethodSims\_getAllVals(

paste0(se\_string, param), solver, reslist)

param\_val <- spec@model$fixed.pars[[param]]

contained <- (param\_val <= est + moe\_est) & (param\_val >= est - moe\_est)

return(mean(contained, na.rm = TRUE))

}

optMethodSims\_getMaxRate <- function(solver, maxlist) {

# Gets how frequently a solver found a maximal log likelihood

#

# Args:

# solver: A string for the solver

# maxlist A list created by optMethodSims\_getBestVals with entries

# containing vectors naming the solvers that maximized the log

# likelihood

#

# Return:

# The proportion of times the solver maximized the log likelihood

maxed <- sapply(maxlist, function(l) {

solver %in% l$solvers

})

return(mean(maxed))

}

optMethodSims\_getFailureRate <- function(solver, reslist) {

# Computes the proportion of times a solver failed to converge.

#

# Args:

# solver: A string for the solver

# reslist: A list created by optMethodSims

#

# Return:

# Numeric proportion of times a solver failed to converge

failed <- sapply(reslist, function(l) {

is.na(l[solver, "LLH"])

})

return(mean(failed))

}

# Vectorization

optMethodSims\_getCaptureRate <- Vectorize(optMethodSims\_getCaptureRate,

vectorize.args = "solver")

optMethodSims\_getMaxRate <- Vectorize(optMethodSims\_getMaxRate,

vectorize.args = "solver")

optMethodSims\_getFailureRate <- Vectorize(optMethodSims\_getFailureRate,

vectorize.args = "solver")

I first create tables containing, for a fixed sample size and model:

* The rate at which a solver attains the highest log likelihood among all solvers for a series
* The rate at which a solver failed to converge
* The rate at which a roughly 95% confidence interval based on the solver’s solution managed to contain the true parameter value for each parameter (referred to as the “capture rate”, and using the robust standard errors)

solver\_table <- function(reslist, tags, spec) {

# Creates a table describing important solver statistics

#

# Args:

# reslist: A list created by optMethodSims

# tags: A vector with strings naming all solvers to include in the table

# spec: A ugarchspec specification with the fixed parameters containing the

# true parameter value

#

# Return:

# A matrix containing metrics describing the performance of the solvers

params <- names(spec1@model$fixed.pars)

max\_rate <- optMethodSims\_getMaxRate(tags, optMethodSims\_getBestVals(reslist))

failure\_rate <- optMethodSims\_getFailureRate(tags, reslist)

capture\_rate <- lapply(params, function(p) {

optMethodSims\_getCaptureRate(p, tags, reslist, spec = spec)

})

return\_mat <- cbind("Maximization Rate" = max\_rate,

"Failure Rate" = failure\_rate)

capture\_mat <- do.call(cbind, capture\_rate)

colnames(capture\_mat) <- paste(params, "95% CI Capture Rate")

return\_mat <- cbind(return\_mat, capture\_mat)

return(return\_mat)

}

**Model 1, n = 100**

as.data.frame(round(solver\_table(spec1\_n100, tags, spec1) \* 100, digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ------------------------- ------------------ ------------- -------------------------- --------------------------- --------------------------

## nlminb 16.2 20.0 21.8 29.2 24.0

## solnp 0.1 0.0 13.7 24.0 15.4

## lbfgs 15.1 35.2 56.6 67.9 58.0

## gosolnp 20.3 0.0 20.3 32.6 21.9

## hybrid 0.1 0.0 13.7 24.0 15.4

## nloptr+COBYLA 0.0 0.0 6.3 82.6 19.8

## nloptr+BOBYQA 0.0 0.0 5.4 82.1 18.5

## nloptr+PRAXIS 15.8 0.0 42.1 54.5 44.1

## nloptr+NELDERMEAD 0.4 0.0 5.7 19.3 8.1

## nloptr+SBPLX 0.1 0.0 7.7 85.7 24.1

## nloptr+AUGLAG+COBYLA 0.0 0.0 6.1 84.5 19.9

## nloptr+AUGLAG+BOBYQA 0.1 0.0 6.5 83.2 19.4

## nloptr+AUGLAG+PRAXIS 22.6 0.0 41.2 54.6 44.1

## nloptr+AUGLAG+NELDERMEAD 11.1 0.0 7.5 18.8 9.7

## nloptr+AUGLAG+SBPLX 0.6 0.0 7.9 86.5 23.0

**Model 1, n = 500**

as.data.frame(round(solver\_table(spec1\_n500, tags, spec1) \* 100, digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ------------------------- ------------------ ------------- -------------------------- --------------------------- --------------------------

## nlminb 21.2 0.4 63.3 67.2 63.8

## solnp 0.1 0.2 32.2 35.6 32.7

## lbfgs 4.5 41.3 85.0 87.6 85.7

## gosolnp 35.1 0.0 69.0 73.2 69.5

## hybrid 0.1 0.0 32.3 35.7 32.8

## nloptr+COBYLA 0.0 0.0 3.2 83.3 17.8

## nloptr+BOBYQA 0.0 0.0 3.5 81.5 18.1

## nloptr+PRAXIS 18.0 0.0 83.9 87.0 84.2

## nloptr+NELDERMEAD 0.0 0.0 16.4 20.7 16.7

## nloptr+SBPLX 0.1 0.0 3.7 91.4 15.7

## nloptr+AUGLAG+COBYLA 0.0 0.0 3.2 83.3 17.8

## nloptr+AUGLAG+BOBYQA 0.0 0.0 3.5 81.5 18.1

## nloptr+AUGLAG+PRAXIS 21.9 0.0 80.2 87.4 83.4

## nloptr+AUGLAG+NELDERMEAD 0.6 0.0 20.0 24.0 20.5

## nloptr+AUGLAG+SBPLX 0.0 0.0 3.7 91.4 15.7

**Model 1, n = 1000**

as.data.frame(round(solver\_table(spec1\_n1000, tags, spec1) \* 100, digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ------------------------- ------------------ ------------- -------------------------- --------------------------- --------------------------

## nlminb 21.5 0.1 88.2 86.1 87.8

## solnp 0.4 0.2 54.9 53.6 54.6

## lbfgs 1.1 44.8 91.5 88.0 91.8

## gosolnp 46.8 0.0 87.2 85.1 87.0

## hybrid 0.5 0.0 55.0 53.6 54.7

## nloptr+COBYLA 0.0 0.0 4.1 74.5 15.0

## nloptr+BOBYQA 0.0 0.0 3.6 74.3 15.9

## nloptr+PRAXIS 17.7 0.0 92.6 90.2 92.2

## nloptr+NELDERMEAD 0.0 0.0 30.5 29.6 30.9

## nloptr+SBPLX 0.0 0.0 3.0 82.3 11.6

## nloptr+AUGLAG+COBYLA 0.0 0.0 4.1 74.5 15.0

## nloptr+AUGLAG+BOBYQA 0.0 0.0 3.6 74.3 15.9

## nloptr+AUGLAG+PRAXIS 13.0 0.0 83.4 93.9 86.7

## nloptr+AUGLAG+NELDERMEAD 0.0 0.0 34.6 33.8 35.0

## nloptr+AUGLAG+SBPLX 0.0 0.0 3.0 82.3 11.6

**Model 2, n = 100**

as.data.frame(round(solver\_table(spec2\_n100, tags, spec2) \* 100, digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ------------------------- ------------------ ------------- -------------------------- --------------------------- --------------------------

## nlminb 8.2 24.2 22.3 34.7 23.9

## solnp 0.3 0.0 21.1 32.6 21.3

## lbfgs 11.6 29.5 74.9 73.2 70.4

## gosolnp 19.0 0.0 31.9 41.2 30.8

## hybrid 0.3 0.0 21.1 32.6 21.3

## nloptr+COBYLA 0.0 0.0 20.5 94.7 61.7

## nloptr+BOBYQA 0.2 0.0 19.3 95.8 62.2

## nloptr+PRAXIS 16.0 0.0 70.2 57.2 52.8

## nloptr+NELDERMEAD 0.2 0.0 7.8 27.8 14.1

## nloptr+SBPLX 0.1 0.0 24.9 91.0 65.0

## nloptr+AUGLAG+COBYLA 0.0 0.0 21.2 95.1 62.5

## nloptr+AUGLAG+BOBYQA 0.9 0.0 20.1 96.2 62.5

## nloptr+AUGLAG+PRAXIS 38.8 0.0 70.4 57.2 52.7

## nloptr+AUGLAG+NELDERMEAD 14.4 0.0 10.7 26.0 16.1

## nloptr+AUGLAG+SBPLX 0.1 0.0 25.8 91.9 65.5

**Model 2, n = 500**

as.data.frame(round(solver\_table(spec2\_n500, tags, spec2) \* 100, digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ------------------------- ------------------ ------------- -------------------------- --------------------------- --------------------------

## nlminb 1.7 1.6 35.0 37.2 34.2

## solnp 0.1 0.2 46.2 48.6 45.3

## lbfgs 2.2 38.4 85.2 88.1 82.3

## gosolnp 5.2 0.0 74.9 77.8 72.7

## hybrid 0.1 0.0 46.1 48.5 45.2

## nloptr+COBYLA 0.0 0.0 8.2 100.0 40.5

## nloptr+BOBYQA 0.0 0.0 9.5 100.0 41.0

## nloptr+PRAXIS 17.0 0.0 83.8 85.1 81.0

## nloptr+NELDERMEAD 0.0 0.0 26.9 38.2 27.0

## nloptr+SBPLX 0.0 0.0 8.2 100.0 40.2

## nloptr+AUGLAG+COBYLA 0.0 0.0 8.2 100.0 40.5

## nloptr+AUGLAG+BOBYQA 0.0 0.0 9.5 100.0 41.0

## nloptr+AUGLAG+PRAXIS 77.8 0.0 84.4 85.4 81.3

## nloptr+AUGLAG+NELDERMEAD 1.1 0.0 32.5 40.3 32.3

## nloptr+AUGLAG+SBPLX 0.0 0.0 8.2 100.0 40.2

**Model 2, n = 1000**

as.data.frame(round(solver\_table(spec2\_n1000, tags, spec2) \* 100, digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ------------------------- ------------------ ------------- -------------------------- --------------------------- --------------------------

## nlminb 2.7 0.7 64.1 68.0 63.8

## solnp 0.0 0.0 70.1 73.8 69.8

## lbfgs 0.0 43.4 90.6 91.5 89.9

## gosolnp 3.2 0.0 87.5 90.3 86.9

## hybrid 0.0 0.0 70.1 73.8 69.8

## nloptr+COBYLA 0.0 0.0 2.3 100.0 20.6

## nloptr+BOBYQA 0.0 0.0 2.5 100.0 22.6

## nloptr+PRAXIS 14.1 0.0 89.1 91.3 88.5

## nloptr+NELDERMEAD 0.0 0.0 46.3 55.6 45.4

## nloptr+SBPLX 0.0 0.0 2.2 100.0 19.5

## nloptr+AUGLAG+COBYLA 0.0 0.0 2.3 100.0 20.6

## nloptr+AUGLAG+BOBYQA 0.0 0.0 2.5 100.0 22.6

## nloptr+AUGLAG+PRAXIS 85.5 0.0 89.1 91.3 88.5

## nloptr+AUGLAG+NELDERMEAD 0.3 0.0 51.9 58.2 51.3

## nloptr+AUGLAG+SBPLX 0.0 0.0 2.2 100.0 19.5

These tables already reveal a lot of information. In general it seems that the AUGLAG-PRAXIS method (the augmented Lagrangian method using the principal axis solver) provided in NLOpt does best for model 2 especially for large sample sizes, while for model 1 the gosolnp method, which uses the solnp solver by Yinyu Ye but with random initializations and restarts, seems to win out for larger sample sizes.

The bigger story, though, is the failure of any method to be the “best”, especially in the case of smaller sample sizes. While there are some optimizers that consistently fail to attain the maximum log-likelihood, no optimizer can claim to consistently obtain the best result. Additionally, different optimizers seem to perform better with different models. The implication for real-world data–where the true model parameters are never known–is to try every optimizer (or at least those that have a chance of maximizing the log-likelihood) and pick the results that yield the largest log-likelihood. No algorithm is trustworthy enough to be the go-to algorithm.

Let’s now look at plots of the estimated distribution of the parameters. First comes a helper function.

library(ggplot2)

solver\_density\_plot <- function(param, tags, list\_reslist, sample\_sizes, spec) {

# Given a parameter, creates a density plot for each solver's distribution

# at different sample sizes

#

# Args:

# param: A string for the parameter to plot

# tags: A character vector containing the solver names

# list\_reslist: A list of lists created by optMethodSimsf, one for each

# sample size

# sample\_sizes: A numeric vector identifying the sample size corresponding

# to each object in the above list

# spec: A ugarchspec object containing the specification that generated the

# datasets

#

# Returns:

# A ggplot object containing the plot generated

p <- spec@model$fixed.pars[[param]]

nlist <- lapply(list\_reslist, function(l) {

optlist <- lapply(tags, function(t) {

return(na.omit(optMethodSims\_getAllVals(param, t, l)))

})

names(optlist) <- tags

df <- stack(optlist)

names(df) <- c("param", "optimizer")

return(df)

})

ndf <- do.call(rbind, nlist)

ndf$n <- rep(sample\_sizes, times = sapply(nlist, nrow))

ggplot(ndf, aes(x = param)) + geom\_density(fill = "black", alpha = 0.5) +

geom\_vline(xintercept = p, color = "blue") +

facet\_grid(optimizer ~ n, scales = "free\_y")

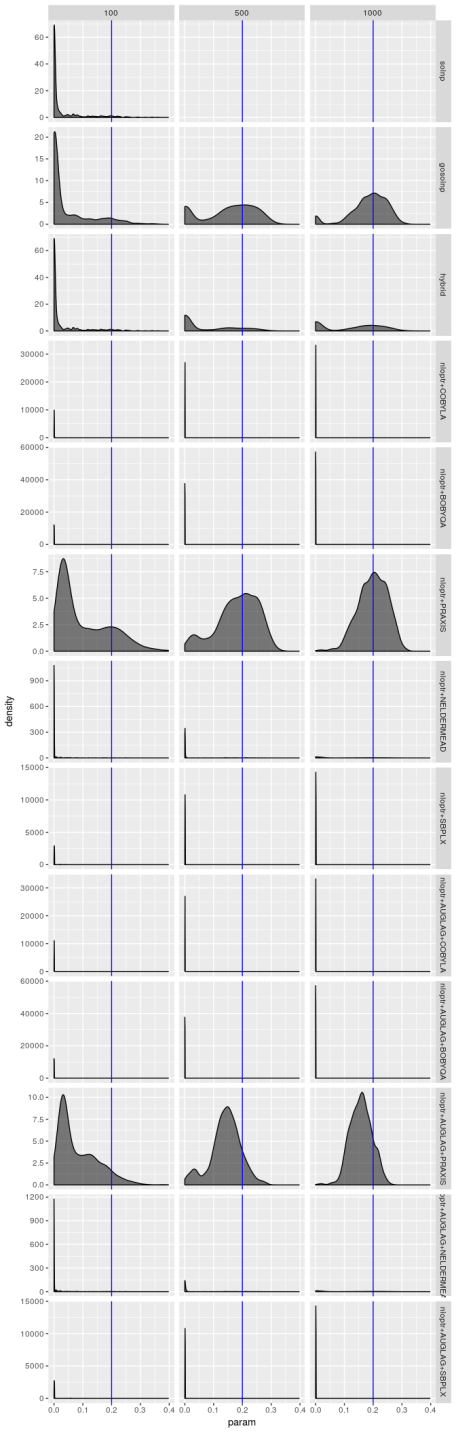
}

Now for plots.

**Estimated \omega, model 1**

solver\_density\_plot("omega", tags, list(spec1\_n100, spec1\_n500, spec1\_n1000),

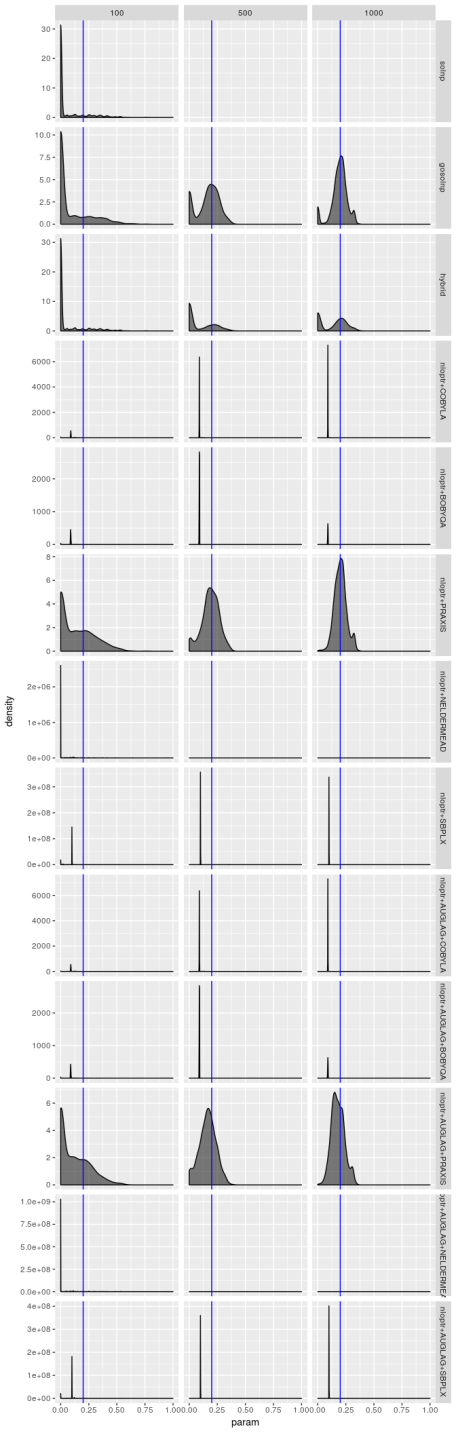
c(100, 500, 1000), spec1)



**Estimated \alpha, model 1**

solver\_density\_plot("alpha1", tags, list(spec1\_n100, spec1\_n500, spec1\_n1000),

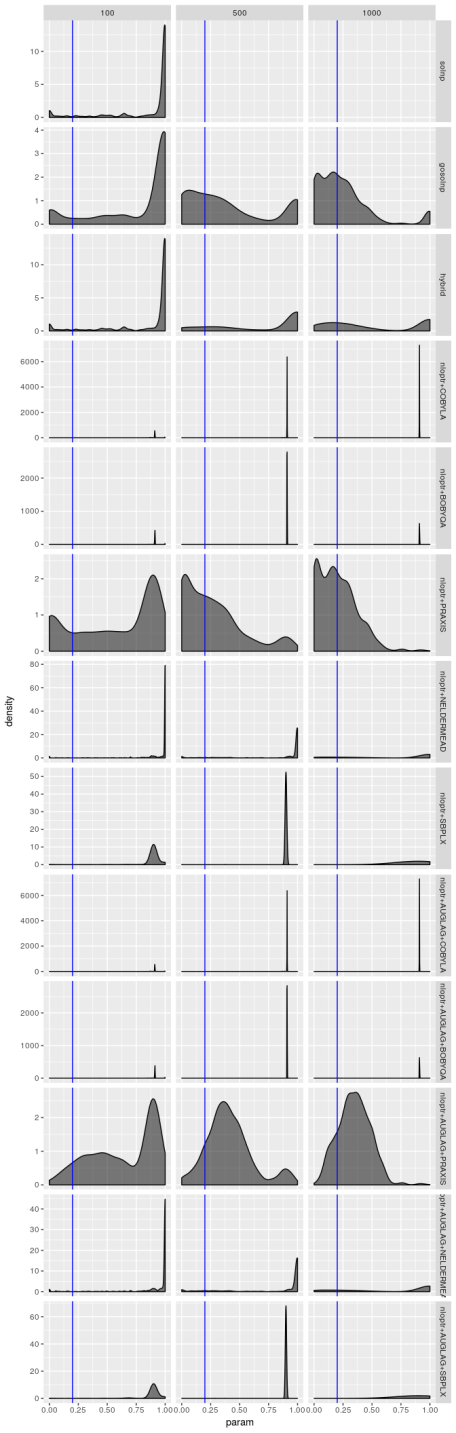
c(100, 500, 1000), spec1)



**Estimated \beta, model 1**

solver\_density\_plot("beta1", tags, list(spec1\_n100, spec1\_n500, spec1\_n1000),

c(100, 500, 1000), spec1)



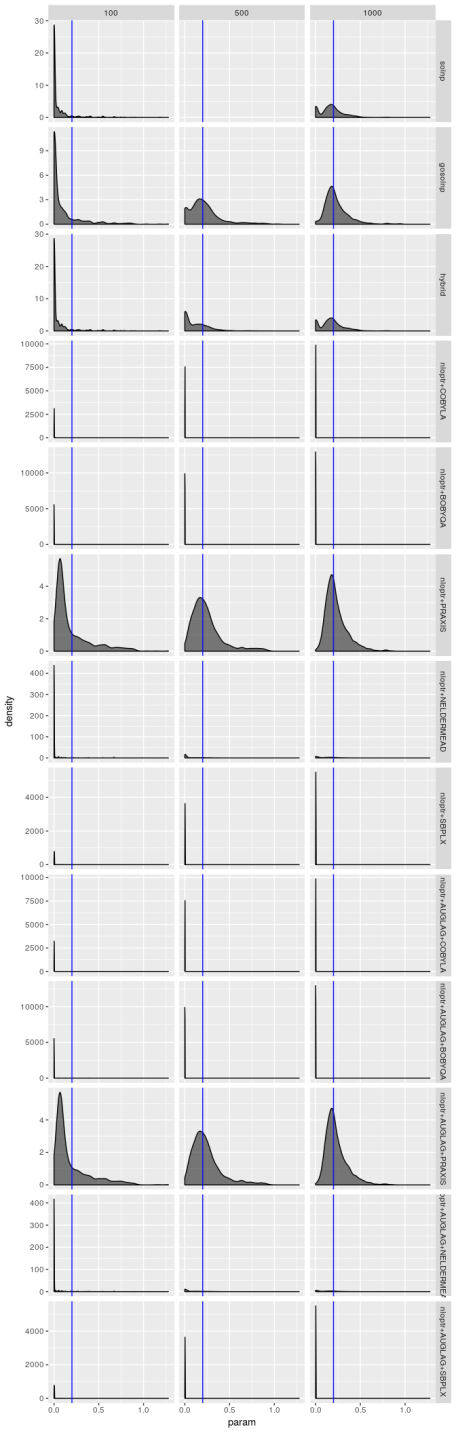
Bear in mind that there are only 1,000 simulated series and the optimizers produce solutions for each series, so in principle optimizer results should not be independent, yet the only time these density plots look the same is when the optimizer performs terribly. But even when an optimizer isn’t performing terribly (as is the case for the gosolnp, PRAXIS, and AUGLAG-PRAXIS methods) there’s evidence of artifacts around 0 for the estimates of \omegaand \alphaand 1 for \beta. These artifacts are more pronounced for smaller sample sizes. That said, for the better optimizers the estimators look almost unbiased, especially for \omegaand \alpha, but their spread is large even for large sample sizes, especially for \beta‘s estimator. That’s not the case for the AUGLAG-PRAXIS optimizer, though; it appears to produce biased estimates.

Let’s look at plots for model 2.

**Estimated \omega, model 2**

solver\_density\_plot("omega", tags, list(spec2\_n100, spec2\_n500, spec2\_n1000),

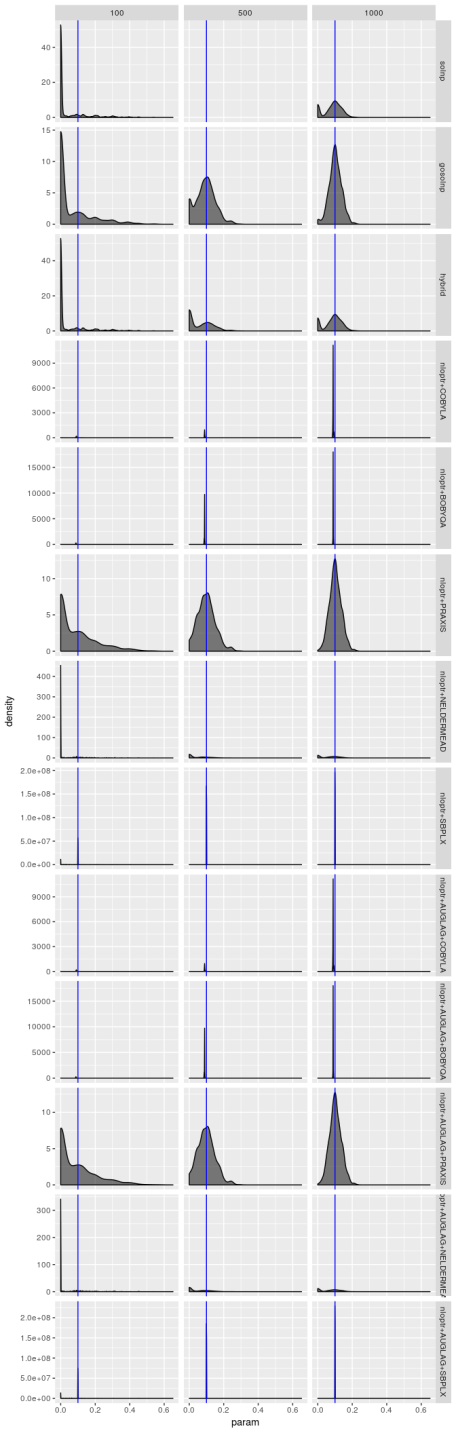
c(100, 500, 1000), spec2)



**Estimated \alpha, model 2**

solver\_density\_plot("alpha1", tags, list(spec2\_n100, spec2\_n500, spec2\_n1000),

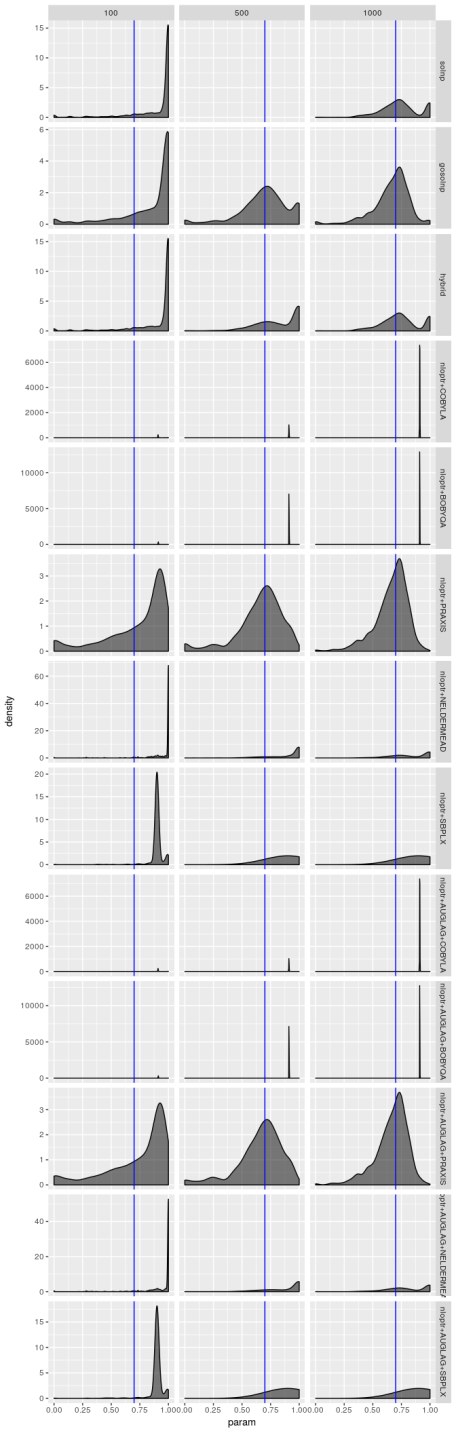
c(100, 500, 1000), spec2)



**Estimated \beta, model 2**

solver\_density\_plot("beta1", tags, list(spec2\_n100, spec2\_n500, spec2\_n1000),

c(100, 500, 1000), spec2)



The estimators don’t struggle as much for model 2, but the picture is still hardly rosy. The PRAXIS and AUGLAG-PRAXIS methods seem to perform well, but far from spectacularly for small sample sizes.

So far, my experiments suggest practitioners should not rely on any one optimizer but instead to try different ones and choose the results that have the largest log-likelihood. Suppose we call this optimization routine the “best” optimizer. how does this optimizer perform?

Let’s find out.

**Model 1, n = 100**

as.data.frame(round(solver\_table(

optMethodSims\_getBestVals(spec1\_n100, reslike = TRUE),

"best", spec1) \* 100,

digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ----- ------------------ ------------- -------------------------- --------------------------- --------------------------

## best 100 0 49.5 63.3 52.2

**Model 1, n = 500**

as.data.frame(round(solver\_table(

optMethodSims\_getBestVals(spec1\_n500, reslike = TRUE),

"best", spec1) \* 100,

digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ----- ------------------ ------------- -------------------------- --------------------------- --------------------------

## best 100 0 86 88.8 86.2

**Model 1, n = 1000**

as.data.frame(round(solver\_table(

optMethodSims\_getBestVals(spec1\_n1000, reslike = TRUE),

"best", spec1) \* 100,

digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ----- ------------------ ------------- -------------------------- --------------------------- --------------------------

## best 100 0 92.8 90.3 92.4

**Model 2, n = 100**

as.data.frame(round(solver\_table(

optMethodSims\_getBestVals(spec2\_n100, reslike = TRUE),

"best", spec2) \* 100,

digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ----- ------------------ ------------- -------------------------- --------------------------- --------------------------

## best 100 0 55.2 63.2 52.2

**Model 2, n = 500**

as.data.frame(round(solver\_table(

optMethodSims\_getBestVals(spec2\_n500, reslike = TRUE),

"best", spec2) \* 100,

digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ----- ------------------ ------------- -------------------------- --------------------------- --------------------------

## best 100 0 83 86.3 80.5

**Model 2, n = 1000**

as.data.frame(round(solver\_table(

optMethodSims\_getBestVals(spec2\_n1000, reslike = TRUE),

"best", spec2) \* 100,

digits = 1))

## Maximization Rate Failure Rate omega 95% CI Capture Rate alpha1 95% CI Capture Rate beta1 95% CI Capture Rate

## ----- ------------------ ------------- -------------------------- --------------------------- --------------------------

## best 100 0 88.7 91.4 88.1

Bear in mind that we evaluate the performance of the “best” optimizer by the CI capture rate, which should be around 95%. The “best” optimizer obviously has good performance but does not outperform all optimizers. This is disappointing; I had hoped that the “best” optimizer would have the highly desirable property of a 95% capture rate. Performance is nowhere near that except for larger sample sizes. Either the standard errors are being underestimated or for small sample sizes the Normal distribution poorly describes the actual distribution of the estimators (which means multiplying by two does not lead to intervals with the desired confidence level).

Interestingly, there is no noticeable difference in performance between the two models for this “best” estimator. This suggests to me that the seemingly better results for models often seen in actual data might be exploiting the bias of the optimizers.

Let’s look at the distribution of the estimated parameters.

**Estimated \omega, model 1**

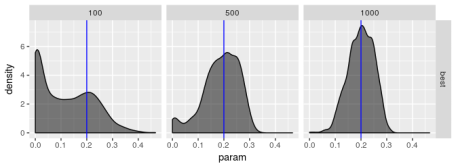
solver\_density\_plot("omega", "best",

lapply(list(spec1\_n100, spec1\_n500, spec1\_n1000),

function(l) {optMethodSims\_getBestVals(l,

reslike = TRUE)}),

c(100, 500, 1000), spec1)



**Estimated \alpha, model 1**

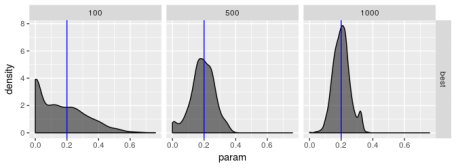
solver\_density\_plot("alpha1", "best",

lapply(list(spec1\_n100, spec1\_n500, spec1\_n1000),

function(l) {optMethodSims\_getBestVals(l,

reslike = TRUE)}),

c(100, 500, 1000), spec1)



**Estimated \beta, model 1**

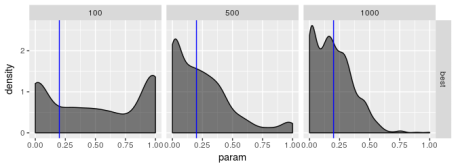
solver\_density\_plot("beta1", "best",

lapply(list(spec1\_n100, spec1\_n500, spec1\_n1000),

function(l) {optMethodSims\_getBestVals(l,

reslike = TRUE)}),

c(100, 500, 1000), spec1)



**Estimated \omega, model 2**

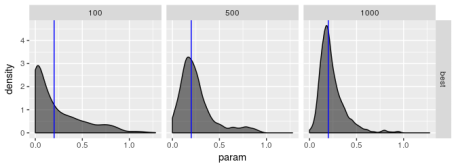
solver\_density\_plot("omega", "best",

lapply(list(spec2\_n100, spec2\_n500, spec2\_n1000),

function(l) {optMethodSims\_getBestVals(l,

reslike = TRUE)}),

c(100, 500, 1000), spec2)



**Estimated \alpha, model 2**

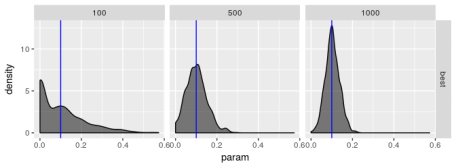
solver\_density\_plot("alpha1", "best",

lapply(list(spec2\_n100, spec2\_n500, spec2\_n1000),

function(l) {optMethodSims\_getBestVals(l,

reslike = TRUE)}),

c(100, 500, 1000), spec2)



**Estimated \beta, model 2**

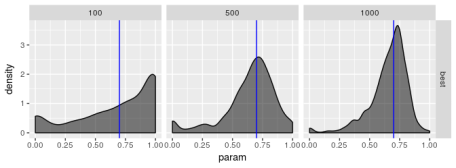
solver\_density\_plot("beta1", "best",

lapply(list(spec2\_n100, spec2\_n500, spec2\_n1000),

function(l) {optMethodSims\_getBestVals(l,

reslike = TRUE)}),

c(100, 500, 1000), spec2)



The plots suggest that the “best” estimator still shows some pathologies even though it behaves less poorly than the other estimators. I don’t see evidence for bias in parameter estimates regardless of choice of model but I’m not convinced the “best” estimator truly maximizes the log-likelihood, especially for smaller sample sizes. the estimates for \betaare especially bad. Even if the standard error for \betashould be large I don’t think it should show the propensity for being zero or one that these plots reveal.

**Conclusion**

The reason for the hang up was because I wanted a literature review of alternative ways to estimate the parameters of a GARCH model. Unfortunately I never completed such a review, and I’ve decided to release this article regardless.

Life happens, though, and I didn’t complete this review. The project moved on and the problem of estimating GARCH model parameters well was essentially avoided. That said, I want to revisit this point, perhaps exploring how techniques such as simulated annealing do for estimating GARCH model parameters.

So for now, if you’re a practitioner, what should you do when estimating a GARCH model? I would say don’t take for granted that the default estimation procedure your package uses will work. You should explore different procedure and different parameter choices and go with the results that lead to the largest log-likelihood value. I showed how this could be done in an automated fashion but you should be prepared to *manually* pick the model with the best fit (as determined by the log-likelihood). If you don’t do this the model you estimated may not actually be the one for which theory works.

I will say it again, one last time, in the last sentence of this article for extra emphasis: *don’t take numerical techniques and results for granted!*

sessionInfo()

## R version 3.4.2 (2017-09-28)

## Platform: i686-pc-linux-gnu (32-bit)

## Running under: Ubuntu 16.04.2 LTS

##

## Matrix products: default

## BLAS: /usr/lib/libblas/libblas.so.3.6.0

## LAPACK: /usr/lib/lapack/liblapack.so.3.6.0

##

## locale:

## [1] LC\_CTYPE=en\_US.UTF-8 LC\_NUMERIC=C

## [3] LC\_TIME=en\_US.UTF-8 LC\_COLLATE=en\_US.UTF-8

## [5] LC\_MONETARY=en\_US.UTF-8 LC\_MESSAGES=en\_US.UTF-8

## [7] LC\_PAPER=en\_US.UTF-8 LC\_NAME=C

## [9] LC\_ADDRESS=C LC\_TELEPHONE=C

## [11] LC\_MEASUREMENT=en\_US.UTF-8 LC\_IDENTIFICATION=C

##

## attached base packages:

## [1] parallel stats graphics grDevices utils datasets methods

## [8] base

##

## other attached packages:

## [1] ggplot2\_2.2.1 rugarch\_1.3-8 printr\_0.1

##

## loaded via a namespace (and not attached):

## [1] digest\_0.6.16 htmltools\_0.3.6

## [3] SkewHyperbolic\_0.3-2 expm\_0.999-2

## [5] scales\_0.5.0 DistributionUtils\_0.5-1

## [7] Rsolnp\_1.16 rprojroot\_1.2

## [9] grid\_3.4.2 stringr\_1.3.1

## [11] knitr\_1.17 numDeriv\_2016.8-1

## [13] GeneralizedHyperbolic\_0.8-1 munsell\_0.4.3

## [15] pillar\_1.3.0 tibble\_1.4.2

## [17] compiler\_3.4.2 highr\_0.6

## [19] lattice\_0.20-35 labeling\_0.3

## [21] Matrix\_1.2-8 KernSmooth\_2.23-15

## [23] plyr\_1.8.4 xts\_0.10-0

## [25] spd\_2.0-1 zoo\_1.8-0

## [27] stringi\_1.2.4 magrittr\_1.5

## [29] reshape2\_1.4.2 rlang\_0.2.2

## [31] rmarkdown\_1.7 evaluate\_0.10.1

## [33] gtable\_0.2.0 colorspace\_1.3-2

## [35] yaml\_2.1.14 tools\_3.4.2

## [37] mclust\_5.4.1 mvtnorm\_1.0-6

## [39] truncnorm\_1.0-7 ks\_1.11.3

## [41] nloptr\_1.0.4 lazyeval\_0.2.1

## [43] crayon\_1.3.4 backports\_1.1.1

## [45] Rcpp\_1.0.0